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(54) Title: A GENE EXPRESSION SYSTEM UTILIZING AN RNA POLYMERASE PREBOUND TO DNA			
(57) Abstract			
<p>The present invention relates to a gene expression system in eukaryotic cells. In particular, it relates to a self-initiating and self-sustaining gene expression system utilizing an RNA polymerase prebound to a DNA construct before the introduction of the complex into cells. This system is capable of functioning in the cell cytoplasm without integration into the genome of host cells, and does not require host cell factors for initiation and maintenance of gene expression. Therefore, the invention has a wide range of applications in <i>in vitro</i> and <i>in vivo</i> gene expression, including gene therapy in resting cells.</p>			

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A GENE EXPRESSION SYSTEM UTILIZING
AN RNA POLYMERASE PREBOUND TO DNA

1. INTRODUCTION

5 The present invention relates to a gene expression system in eukaryotic cells. In particular, it relates to a self-initiating and self-sustaining gene expression system utilizing an RNA polymerase prebound to a DNA construct before the introduction of 10 the complex into cells. This system is capable of functioning in the cell cytoplasm without integration into the genome of host cells, and does not require host cell factors for initiation and maintenance of 15 gene expression. Therefore, the invention has a wide range of applications in in vitro and in vivo gene expression, including gene therapy in resting cells.

2. BACKGROUND OF THE INVENTION

20 2.1. EXOGENOUS GENE EXPRESSION
Advances in recombinant DNA technology have permitted the expression of exogenous foreign genes in eukaryotic cells. Foreign gene expression has not only facilitated studies of gene function and the 25 production of properly processed gene products, it has also led to the possibility of correcting gene defects by introducing functionally active genes in gene therapy.

30 A number of gene delivery and expression systems have been developed and successfully used to express foreign genes in eukaryotic cells, and less satisfactorily, in animals. These systems consist primarily of a DNA delivery method and a DNA expression vector. DNA delivery methods include 35 calcium phosphate precipitation (Wigler et al., 1979,

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Proc. Natl. Acad. Sci. USA 76:1373-1376), DEAE-dextran (Sompagnac et al., 1981, Proc. Natl. Acad. Sci. USA 78:7575-7578), lipofection (Felgner et al., 1987, Proc. Natl. Acad. Sci. USA 84:7413-7417),
5 electroporation (Neumann et al., 1982, EMBO J. 7:841-845), retroviruses (Schimotohono et al., 1981, Cell 26:67-77), direct DNA injection (Benvenisty et al., 1986, Proc. Natl. Acad. Sci. USA 83:9551-9555; Wolff et al., 1990, Science 247:1465-1468), specific
10 receptor-mediated DNA uptake (Wu et al., 1988, J. Biol. Chem. 263:14621-14624; Wu et al., 1989, J. Biol. Chem. 264:16985-16987), and more recently, aerosol DNA delivery (Stribling et al., 1992, Proc. Natl. Acad. Sci. USA 89:11277-11281). Expression vectors are in
15 general plasmid DNA containing a gene of interest linked to a regulatory sequence which controls the expression of the gene upon its introduction into a given cell.

Since these gene expression systems usually
20 utilize eukaryotic or mammalian promoters to direct gene transcription, they require entry into host cell nuclei and integration of the exogenous DNA for expression. However, only a very small percentage of the cells which take up DNA contain the foreign DNA
25 within the nuclei of the cells. This problem becomes even more pronounced when whole animals are the targets of foreign gene expression because cell division, which is required for nuclear deposition of the introduced DNA, does not always occur at as high a
30 frequency in the cells of tissues as it does in cultured cell lines, making it difficult for exogenous DNA to integrate into non-dividing cells.

On the other hand, evidence has indicated that the cytoplasmic uptake of foreign DNA by cells
35 and animals is efficient (Felgner et al., 1987, Proc.

Natl. Acad. Sci. USA 84:7413-7417). And, in certain circumstances, the DNA introduced into the cells of whole animal tissues has been observed to remain in the cytoplasm up to several months (Wolff et al., 5 1990, Science 247:1465-1468).

2.2. BACTERIOPHAGE RNA POLYMERASE

Bacteriophage T7 RNA polymerase (RNAP) has been studied extensively in vitro and in vivo in 10 bacteria and eukaryotic cells due to several of its unique biochemical characteristics. In particular, T7 RNAP is a single polypeptide enzyme capable of carrying out transcription with high promoter specificity and efficiency, without the involvement of 15 any other cellular transcription factors (Davanloo et al., 1984, Proc. Natl. Acad. Sci. USA 81:2035-2039; Dunn et al., 1983, J. Mol. Biol. 166:477-535; Studier et al., 1990, Methods Enzymol. 185:60-89). For example, recombinant vaccinia viruses containing a T7 20 RNAP gene or a cell line which constitutively expresses T7 RNAP have been used to promote expression of genes of interest linked to T7 promoter in the cytoplasm of mammalian cells (Fuerst et al., 1986, Proc. Natl. Acad. Sci. 83:8122-8126; Fuerst et al., 25 1989, J. Mol. Biol. 206:333-348; Elroy-Stein et al., 1989, Proc. Natl. Acad. Sci. USA 86:6126-6130; Elroy-Stein et al., 1990, Proc. Natl. Acad. Sci. USA 87: 6743-6747). When a chloramphenicol 30 acetyltransferase (CAT) gene was inserted into a T7 promoter-containing mammalian vector and transfected into a stable cell line which also expressed T7 RNAP, as high as 30% of cytoplasmic proteins was found to be the CAT enzymes in the transfected cells (Elroy-Stein et al., 1990, Proc. Natl. Acad. Sci. USA 87: 35 6743-6747). However, a major limitation of such a T7

gene expression system is that it must be performed in cell lines which also express the T7 RNAP gene, which is a bacterial gene not expressed endogenously in eukaryotic cells.

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3. SUMMARY OF THE INVENTION

The present invention relates to a self-initiating, self-sustaining gene expression system in eukaryotic cells utilizing an RNA polymerase 10 prebound to a DNA molecule, and its use in the expression of genes in in vitro cell lines and in in vivo tissues, including gene therapy. The unique components of this expression system allow gene transcription without requiring the involvement of 15 host cell factors and without integrating into the host cell genome.

The present invention is based, in part, on Applicants' discovery of significant gene expression from a DNA construct complexed to T7 RNAP, which 20 construct contains a T7 RNAP gene driven by a T7 promoter, and another nucleotide sequence encoding a functional or a reporter gene i.e. a gene of interest, under the control of a second T7 promoter (T7T7/T7-gene construct). One unique feature of the 25 construct which distinguishes this system from other gene expression systems is that both the initiation and maintenance of gene expression depend upon the binding of T7 RNAP to DNA prior to the introduction of the construct into host cells. The complex of 30 prebound RNAP to plasmid DNA is stable without detachment during entry into cells. Once the DNA-RNAP enzyme complex enters the cytoplasm of the cells, transcription is initiated immediately by the prebound T7 RNAP via the prebound T7 promoters in the plasmid. 35 The subsequent production of T7 RNAP enzyme, in turn,

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triggers transcription of the functional/reporter gene as well as continued synthesis of additional T7 RNAP, thus it is both a self-initiating and a self-sustaining system. The transcription of both the 5 T7 RNAP and the functional/reporter genes can be driven repeatedly by newly synthesized T7 RNAP in the cell cytoplasm without nuclear integration.

The human growth hormone (hGH) gene encoding a secretory protein, the chloramphenicol 10 acetyltransferase (CAT) gene and the luciferase gene, both encoding intracellular enzymes have been expressed using the expression system of the invention in mouse and human cell lines, and in different mouse tissues in vivo. The expression system described 15 herein provides a method for the efficient expression of cytoplasmically introduced genetic sequences encoding proteins, antisense RNA or ribozymes in a wide variety of host cells, including mitotically quiescent cells.

20

4. BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1. Schematic presentation of self-initiation and positive feedback loop features of the T7T7/T7-gene expression system. A.

25 Cotransfected, prebound T7 RNAP initiate transcription from T7 promoters (P_{T7}) on both T7T7 and T7hGH sequences (1 and 1'). EMC serves as cap independent sequence in translation for the T7 and hGH transcripts; 30 B. newly synthesized T7 RNAPs replenish the polymerase pool in the cytoplasm of the transfected cells (2 and 2'); and C. maintenance of the T7 and hGH gene expression in the cells (3 and 3'). In

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this plasmid, hGH cDNA can be replaced to express other cDNA.

Figure 2. Construction of pT7T7 plasmid. pTM-1 and pAR1173 were used as starting plasmids. pTM-1 contained a bacteriophage T7 ($\phi 10$) promoter (P_{T7}), a capping independent (EMC) sequence which facilitated translation of uncapped transcripts, as well as a T7 transcription termination sequence (T_{T7}). Polycloning site (PCS) in pTM-1 contained several restriction sites for cDNA insertion. pAR1173 provided a T7 RNAP gene (Bam HI fragment). In pTLO-1, Lac I and Op represented the Lac I gene and the operator sequence, respectively.

Figure 3. Structure of plasmids pT7hGH and pT7CAT. A. pT7hGH, B. PT7CAT. pT7hGH and pT7CAT were constructed using similar strategies. Both genes were individually inserted into pTM-1. NcoI site (5'-proximal one for PT7CAT) in both plasmids represented the junction of the EMC sequence and the inserted genes (hGH or CAT). ATG codon in the NcoI site represented the first amino acid residue for both hGH and CAT proteins. Internal restriction sites Bgl II for the hGH cDNA and NcoI for the CAT gene were used for restriction analysis to identify the inserted genes. pT7hGH and pT7CAT did not contain either Lac I or operator sequences.

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Figure 4. Construction of pT7T7/T7hGH. Smaller ClaI/EagI fragment from PT7hGH was isolated and inserted into EagI site of pT7T7 by a blunt end ligation. The orientation of the 5 T7hGH fragment in pT7T7/T7hGH was the same as that of the T7T7 sequence. In this plasmid, T7hGH sequence had a structural arrangement similar to T7T7 except that the operator was absent from T7hGH sequence.

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Figure 5. Expression of hGH in mouse L cells transfected by T7hGH system. Mouse L cells were transfected by either pT7T7 + pT7hGH (dual plasmids) or pT7T7/T7hGH (single plasmid) with prebound T7 RNAP as follows: 15 One μ g of plasmid DNA (0.5 μ g + 0.5 μ g for dual plasmids) was coincubated with 125 U of T7 RNAP at room temperature for 10 minutes, 20 μ l of lipofectin was mixed with the DNA-enzyme complexes to a final volume of 100 μ l. Following 5 min room 20 temperature incubation, the liposome-DNA-enzyme mixture was added to mouse L cells in a 6-well cell culture dish. Cell samples transfected by pT7T7/T7hGH alone, 25 pT7hGH+T7 RNAP, or pMThGH (1 μ g DNA/well) served as negative and positive controls, respectively. Each point in curves represented an average value of at least three individual measurements. Error bars 30 represent standard deviations of the measurements. ■ pT7T7/T7hGH alone, □ pT7hGH+T7 RNAP, Δ pMThGH, 35 ○ pT7T7+pT7hGH+T7 RNAP, and pT7T7/T7hGH+T7 RNAP.

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Figure 6. Expression of the CAT gene in mouse L cells transfected with pT7T7 + pT7CAT with prebound T7 RNAP. Transfection was performed the same way as described. 24 hours after the transfection, the cells were harvested and assayed for CAT activity. Cell samples transfected with either pT7CAT alone or pT7CAT with prebound T7 RNAP served as controls. Error bars represent standard deviations of the measurements.

Figure 7A and 7B.

Northern analysis of the T7 and hGH mRNA expressed by mouse L cells transfected by the T7 system. The cells were transfected by the T7 system, and total RNA was isolated. Following gel electrophoresis and transfer of resolved RNA on a membrane, the membrane was hybridized first with a 2.6 kbp ³²P-labeled T7 probe, then by a 0.9 kbp hGH probe after stripping off the first T7 probe. Actin mRNA in each sample served as a RNA concentration reference. 7A. Northern blot using T7 probe. A=cells transfected by pT7T7/T7hGH, B=pT7hGH+T7 RNAP, A and B were isolated 24 hr after the transfection. C and D=pT7T7/T7hGH+T7 RNAP; sample C was isolated 24 hr post transfection, D=48 hr. 7B. Same membrane hybridized with a hGH probe. Lane order is the same as in 7A.

Figure 8. Luciferase expression in mouse tail connective tissues by injection of pT7T7/T7Luc plasmid DNA. Different amounts

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of plasmid DNA pT7T7/T7Luc, were complexed with T7 RNAP in test tubes, and injected into the tail connective tissues BALB/c mice of 1-2 months of age. Twenty four hours post injection, injected tail tissues were homogenized using 300 μ l Luciferase Lysis Buffer, followed by centrifugation to precipitate cell debris. Five μ l of the cell lysate from each sample was used for luciferase assays.

Figure 9. Luciferase expression in mouse leg muscles by injection of pT7T7/T7Luc plasmid DNA. One hundred μ l of solution, which contained either PBS, or 25 μ g pT7T7/T7Luc complexed with T7 RNAP, were injected into mouse hind leg muscles using 25G needles. Twenty four hours after injection, the injected muscles were isolated, homogenized, and assayed for luciferase activity.

Figure 10. Luciferase expression in mouse livers by direct DNA injection. One hundred μ l of DMEM solution, which contained 50 or 100 μ g of pT7T7/T7Luc DNA complexed with T7 RNAP, were injected into one lobe of BALB/c mouse livers. Twenty hours after the injection, mouse livers were removed, homogenized, and assayed for luciferase activity.

Figure 11. Luciferase expression in mouse brains by injection of pT7T7/T7Luc plasmid DNA. Fifty μ l of solution, which contained either PBS or pT7T7/T7Luc DNA-T7 RNAP complex, were injected into the brain

tissues of BALB/c mice of 20 days of age. Sixteen hours after the injection, brain tissues were removed and lysed into 400 μ l of Luciferase Lysis Buffer. Ten μ l of cell lysates were assayed for luciferase activity as described previously.

Figure 12. Human growth hormone expression in mouse
brain tissues and in cerebrospinal fluids.
10 Fifty to 80 μ l of solution, which contained
either PBS, pT7T7/T7hGH + T7 RNAP, or
pT7T7/T7hGH + T7 RNAP + lipofectin, were
injected into the cerebrospinal fluid from
the top of the head of mice of 20 days of
15 age. Twenty hours after the injection,
cerebrospinal fluids and brain tissues from
the injected mice were isolated,
homogenized, and assayed for hGH activity
by an hGH RIA. CSF = cerebrospinal fluid,
20 BT = brain tissue.

5. DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to a gene expression system in eukaryotic host cells. This system is different from other expression systems because it relies on the binding of an RNA polymerase to a DNA molecule constructed to contain its cognate promoter and the RNAP coding sequence, and a functional/reporter gene also driven by the same cognate promoter, prior to cell transfection (FIG. 1). This arrangement creates a potential positive feedback loop for the expression of the RNAP gene. Once this DNA-enzyme complex is introduced into eukaryotic cells, the transcription of the RNAP and the functional/reporter genes is initiated by the prebound

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RNAP. The prebound RNAP, which is responsible for the initiation of the expression of both the RNAP and the functional/reporter genes, is replenished by translation of newly synthesized RNAP. This 5 expression system has been designed in such a manner that the expression of the functional/reporter genes can occur in the cytoplasm and does not require any nuclear involvement.

The invention is discussed in more detail in 10 the subsections below, solely for purposes of description and not by way of limitation. For clarity of discussion, the specific procedures and methods described herein are exemplified using specific genes and cell lines; they are merely illustrative for the 15 practice of the invention. Analogous procedures and techniques are equally applicable to all genes and tissues.

5.1. ISOLATION OF A PROTEIN CODING SEQUENCE

20 The expression system of the invention may be used to express any gene of interest in eukaryotic cells. An isolated gene sequence may be inserted into a plasmid in a manner similar to that described in Section 6.1.3., infra. For the isolation of a gene 25 sequence encoding a gene product of interest, messenger RNA (mRNA) may be obtained from cell sources that produce the protein, whereas its genomic sequences may be obtained from any cell source. Genetically-engineered microorganisms or cell lines 30 containing the protein coding sequence may be used as a convenient source of DNA for this purpose.

Either cDNA or genomic libraries may be prepared from the DNA fragments generated using techniques well known in the art. The fragments which 35 encode a specific protein may be identified by

screening such libraries with a nucleotide probe homologous to a portion of the protein which is based on amino acid sequence information obtained from the purified protein. Alternatively, an antibody probe 5 may be used to screen a library generated by expression cloning methods such as λgt11 (Young and Davis, Proc. Natl. Acad. Sci. U.S.A. 80:1194-1198). Where a partial sequence of the gene of interest is known, PCR (polymerase chain reaction), LCR (ligase 10 chain reaction) or the like can be utilized to generate and identify clones. (Innis et al., 1990, PCR Protocols, Academic Press Inc., New York) Although portions of the coding sequence may be utilized for cloning and expression, full length clones, i.e., those 15 containing the entire coding region for a protein, may be preferable for expression. To these ends, techniques well known to those skilled in the art for the isolation of DNA, generation of appropriate restriction fragments, construction of clones and 20 libraries, and screening recombinants may be used. (See, for example, the techniques described in Maniatis et al., 1989, Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory, N.Y.)

Regardless of the method chosen to identify 25 and clone a specific coding sequence, expression cloning methods may be utilized to substantially reduce the screening effort. Recently, a one step procedure for cloning and expressing antibody genes has been reported (McCafferty et al., 1990, Nature 30 348:552-554; Winter and Milstein, 1991, Nature 349:293-299). Based on this technology, any gene may likewise be cloned directly into a vector at a site adjacent to the coat protein gene of a bacteriophage such as λ or fd. The phage carrying the gene of 35 interest expresses the fusion protein on its surface

so that columns containing a specific antibody can be used to select and isolate phage particles with binding activity. A commercially available expression cloning system utilizing Lambda-Zap-bluescript may

5 also be used for the cloning and antibody screening of cDNA libraries (Stratagene, La Jolla, CA). Transient gene expression systems may be utilized to identify the correct gene of interest. For example, the COS cell system (e.g., Gerard & Gluzman, 1986, Mol. Cell.

10 Biol. 6(12) 4570-4577) may be used.

Due to the degeneracy of the nucleotide coding sequences, other DNA sequences which encode analogous amino acid sequences for any known gene may be used in the practice of the present invention for

15 the cloning and expression of its gene product. Such alterations include deletions, additions or substitutions of different nucleotide residues resulting in a sequence that encodes the same or a functionally equivalent gene product. The gene

20 product may contain deletions, additions or substitutions of amino acid residues within the sequence, which result in a silent change thus producing a bioactive product. Such amino acid substitutions may be made on the basis of similarity

25 in polarity, charge, solubility, hydrophobicity, hydrophilicity and/or the amphipathic nature of the residues involved. For example, negatively charged amino acids include aspartic acid and glutamic acid; positively charged amino acids include lysine and

30 arginine; amino acids with uncharged polar head groups having similar hydrophilicity values include the following: leucine, isoleucine, valine; glycine, alanine; asparagine, glutamine; serine, threonine; phenylalanine, tyrosine. Oligonucleotide sequences

35 can be used to increase the copy number of a unique

gene sequence by the polymerase chain reaction. This approach would provide for more specific probes than that obtained using degenerate oligonucleotides.

In addition, other DNA sequences containing 5 structural modifications but without substantial alteration of the biologic activities of the encoded protein may also be used in the practice of the invention. Such modifications include but are limited to additions, deletions or substitutions of amino acid 10 residues in a protein to create additional processing sites and/or elimination of glycosylation sites. For example, the removal of N-linked glycosylation sites in certain proteins results in reduced glycosylation on the expressed products which are particularly 15 useful in yeast expression systems.

5.2. ANTI-SENSE RNA AND RIBOZYMES

Also within the scope of the invention is the expression of oligo-ribonucleotide sequences, that 20 include anti-sense RNA and ribozymes that function to inhibit the translation of a variety of mRNA. Anti-sense RNA acts to directly block the translation of mRNA by binding to targeted mRNA and preventing protein translation, either by inhibition of ribosome 25 binding and/or translocation or by bringing about the nucleic acid degradation of the mRNA molecule itself.

Ribozymes are enzymatic RNA molecules capable of catalyzing the specific cleavage of RNA. The mechanism of ribozyme action involves sequence 30 specific hybridization of the ribozyme molecule to complementary target RNA, followed by a endonucleolytic cleavage. Within the scope of the invention are engineered hammerhead motif ribozyme molecules that specifically and efficiently catalyze 35 endonucleolytic cleavage of mRNA sequences.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites which include the following sequences, GUA, GUU 5 and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides corresponding to the region of the target gene containing the cleavage site may be evaluated for predicted structural features such as secondary structure that may render the oligo- 10 nucleotide sequence unsuitable. The suitability of candidate targets may also be evaluated by testing their accessibility to hybridization with complimentary oligonucleotides, using ribonuclease protection assays.

15 Both anti-sense RNA and ribozymes may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligodeoxyribonucleotides well known in the art such as for 20 example solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by in vitro and in vivo transcription of DNA sequences encoding the antisense RNA molecule. Such 25 DNA sequences may be incorporated into the expression system of the invention.

Various modifications to the DNA molecules may be introduced as a means of increasing intracellular stability and half-life. Possible modifications include but are not limited to the 30 addition of flanking sequences of ribo- or deoxy- nucleotides to the 5' and/or 3' ends of the molecule or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the oligodeoxyribonucleotide backbone.

5.3. SELF-INITIATING AND SELF-SUSTAINING GENE EXPRESSION SYSTEMS

According to one aspect of the present invention, a self-initiating and self-sustaining gene expression system may be constructed by binding a RNA polymerase to a DNA construct containing a cognate promoter of the RNA polymerase, operably linked to a nucleotide sequence encoding the RNA polymerase. Operably linked is used herein to describe the transcription from the promoter by the RNA polymerase in the host cell to produce functional RNA transcripts that can be translated to the RNA polymerase protein. The gene construct that operably links the RNA polymerase encoding sequence to a promoter sequence recognized by the same RNA polymerase is referred to hereinafter as an autogene (Dubendorff and Studier, 1991, J. Mol. Biol. 219:61-68, for the origin of the "autogene" term and an example of a T7 RNA polymerase autogene).

The RNA polymerase of the present invention should be capable of carrying out transcription in the intended eukaryotic host cell. It is preferred that the RNA polymerase is not complex in composition, e.g., composed of multiple subunits, and should have sufficiently high affinity for its cognate promoter so that the complex is maintained during introduction into host cells. A functional RNA polymerase made up of no more than two subunits is preferable. More preferable is a RNA polymerase composed of only homomers. Most preferable is a RNA polymerase that is a small single unit enzyme, that does not require host cell factors for activity, that recognizes its cognate promoter sequence with a high degree of specificity, and that is highly active. Examples of RNA polymerases suitable for use in the present invention include, but are not limited to, the RNA polymerases

of the T7, T3, SP6, or K11 bacteriophages or of the RNA polymerases of mitochondria. (See Chamberlin and Ryan, 1983; Dietz et al., 1990, Mol. Gen. Genet. 221:283-286; Schinkel and Tabak, 1989, Trend Genet. 5 5:149-54).

An autogene construct of the present invention generally includes five components linked in the following 5' to 3' orientation: a cognate promoter of the RNA polymerase, a sequence encoding an 5' 10 untranslated RNA (UTR), a sequence encoding the RNA polymerase, a sequence encoding a 3'-UTR, a cognate transcription terminator of the RNA polymerase. These parts are linked such that transcription by the RNA polymerase from the cognate promoter would initiate at 15 the 5'-end of the 5'-UTR sequence, proceed through the sequences encoding the 5'-UTR, the RNA polymerase and the 3'-UTR sequences, and terminate in the cognate transcription terminator. A transcript produced from such a autogene construct in the eukaryotic host cell 20 would contain a RNA that has the 5'-UTR sequence at its 5'-end, the RNA polymerase sequence in its middle, and the 3'-UTR sequence at its 3'-end.

The cognate promoter sequence can be a promoter sequence cloned from the genome encoding the 25 RNA polymerase. The location of a promoter can be identified by approaches and methods well known in the art, including DNase foot printing of the RNAP bound genome DNA, mutational analysis, etc. Alternatively, where the sequence of a cognate promoter is known, the 30 promoter sequence can be synthesized. It may be advantageous to modify the cognate promoter by attaching at its 3'-end a bacterial operator sequence such as the lac operator. The operator sequence may serve to reduce the toxicity of constructs containing 35 the autogene in the bacteria host. It would be

advantageous that the operator is a short sequence. Preferably the operator is no longer than 50 basepairs. Most preferably the operator is no longer than 25 basepairs.

5 The cognate promoter, unmodified or modified, is operably linked at its 3'-end to the 5'-end of the 5'-UTR encoding sequence such that the transcription by the RNA polymerase from the cognate promoter would initiate RNA synthesis at the beginning 10 (e.g. 5'-end) of the 5'-UTR sequence.

The 5'-UTR sequence encodes a RNA that can enable the formation of translational initiation complex in the eukaryotic host cell without the need for the RNA to be "capped" at it's 5'-end. (Kozak, 15 1983, *Microbiol. Rev.*, 47:1-45). Preferably, the 5'-UTR sequence encodes a RNA sequence from the 5' untranslated region of a picornavirus, such as encephalomyocarditis (EMC) virus, polio virus, mengo virus, etc. Most preferably, the 5'-UTR sequence 20 encodes the 5' untranslated region of the EMC virus RNA (see Elroy-Stein et al., 1989, *Proc. Nat. Acad. Sci. USA* 86:6126-6130). In order to reduce the toxicity of autogene constructs in bacterial host, it may be advantageous to delete or mutate any fortuitous 25 "Shine and Delgarno" sequences present at the 3'-end of the 5'-UTR sequence.

According to the present invention, the 3'-end of the 5'-UTR sequence is linked with the 5'-end of the RNA polymerase-encoding sequence. The RNA 30 polymerase encoding sequence may be obtained by cloning from an appropriate genome. Approaches and methods for cloning RNA polymerase encoding sequences are well known in the art. They include the use of nucleotide probes derived from purified peptides of 35 the RNAP or nucleotide probes from related RNAP to

probe genomic or cDNA libraries or to generate clones using PCR, LCR etc. techniques. Alternatively, where the amino acid or nucleotide sequence of a desired RNA polymerase is known, the RNA polymerase encoding sequence 5 may be synthesized according to the genetic code, using the codon choices most preferred by the host cell which is to express the autogene.

It may be advantageous, where appropriate, to modify the 5'-end of the RNA polymerase encoding 10 sequence to prevent any out-of-frame (relative to the RNA polymerase coding sequence) AUG codons from being formed as a result of linking the polymerase encoding sequence to the 3'-end of the 5'-UTR sequence. The necessary modifications may involve methods, well known 15 in the art including site-specific mutagenesis, addition or deletion of short sequences, and/or any other type of sequence manipulations.

According to the present invention, the 5'-end of the 3'-UTR encoding sequence is linked to the 3'-end of 20 the sequence encoding the RNA polymerase. The 3'-UTR encoding sequence should encode a RNA sequence that would stabilize the autogene transcript in the eukaryotic host cell. Sequences that could serve such a function include those encoding the 3' untranslated 25 RNA sequences of histone genes, or a polyadenylate nucleotide tract, commonly known as polyA tracts. Where a histone 3' untranslated RNA sequence is used as the 3'-UTR of an autogene construct, it is preferable that the source of the histone gene 30 sequence be the eukaryotic host cell which is to express the autogene. Where a polyA encoding sequence is used as the 3'-UTR sequence in the autogene construct, the sequence can be synthesized. The length of the polyA tract encoded by the 3'-UTR 35 sequence preferably is longer than 25 nucleotides and

shorter than 250 nucleotides. A sequence encoding a polyA tract between 40 to 50 nucleotides in length is most preferred.

According to the present invention, the

5 cognate transcription terminator sequence should be attached immediately 3' of the sequence encoding the 3'-UTR. The terminator sequence can be cloned from the genome encoding the RNA polymerase. The approaches and methods for cloning such terminator

10 sequences are well known in the art and include physical mapping the 3' end of RNA transcripts by sequencing, RNase protection, etc. or genetic mapping by mutagenesis, deletion analysis, etc.

15 Alternatively, where the sequence of a cognate terminator is known, the sequence can be synthesized.

In various embodiments of the present invention the RNA polymerase may be composed of several different proteins. In such instances, the present invention may be practiced by constructing an

20 autogene encoding each of the heteromeric proteins, prebinding RNA polymerase to each autogene construct, and introducing simultaneously the protein-DNA complex for every subunit of the RNA polymerase into the host cell. Where the RNA polymerase is composed of few

25 subunits, it may be advantageous to have a single DNA construct contain all the RNA polymerase autogenes.

In another aspect of the present invention, the self-initiating and self-sustaining gene expression system may be used to express other gene

30 products including proteins, anti-sense molecules or ribozymes in eukaryotic host cells. Such gene products may be expressed by introducing into the eukaryotic host cell DNA containing gene constructs encoding the desired gene products together with DNA

35 encoding the RNA polymerase autogene(s) prebound with

the RNA polymerase, in which the gene construct encoding a desired gene product is under the control of a cognate promoter of the RNA polymerase.

According to the present invention, the 5 construction of the gene construct encoding a desired gene product is identical to that of an autogene, except the sequence encoding the desired gene product replaces that encoding the RNA polymerase. That is the gene construct comprises of the same five parts 10 linked in the same 5' to 3' order: a cognate promoter of the RNA polymerase, a sequence encoding a 5'-UTR, a sequence encoding the desired gene product, a sequence encoding a 3'-UTR, a cognate transcription terminator of the RNA polymerase. Where the desired gene product 15 is an antisense RNA or ribozyme, the DNA sequences encoding the 5'-UTR and 3'-UTR may be advantageously omitted from the gene construct.

In addition, a single DNA construct containing both an autogene and a gene of interest 20 under the control of two copies of the same cognate promoter is also within the scope of the invention. A single unit expression system may be more efficient for introduction into host cells than a dual system composed of two separate plasmids.

25 A plasmid containing a gene construct of the present invention may include a selectable marker for propagation of the construct in bacteria. Such selectable markers may contain an antibiotic resistance gene, such as those that confer resistance 30 to ampicillin, kanamycin, tetracycline, streptomycin, or chloramphenicol, and the like. Further, a plasmid containing an autogene construct of the present invention may encode a bacterial expressible repressor gene, wherein the encoded repressor recognizes the 35 operator sequence modifying the RNA polymerase

promoter of the autogene. Examples of applicable repressor genes include those of the lactose, histidine, tryptophane operons, and the like.

A unique feature of the present invention is 5 the ability to prebind RNAP to DNA prior to transfection. An RNAP having an affinity for DNA sufficient to allow its binding during entry into cells may be used for the practice of the invention. In fact, any DNA binding protein with such affinities 10 may be used for this aspect of the invention. Conventional techniques, e.g. calcium phosphate precipitation, lipofection, electroporation, DEAE-transfection, and the like may be used to introduce the complex into host cells.

15

5.4. USES OF SELF-INITIATING AND SELF-SUSTAINING GENE EXPRESSION SYSTEMS

The present invention relates to a gene expression system possessing a number of unique 20 characteristics. This system contains all necessary components for gene expression without the involvement of host cell factors, thus it is both self-initiating and self-sustaining. As a result, this system may provide for cell-type independent gene expression, as 25 evidenced by the comparable expression efficiency of several genes in both human and mouse cells (see Example 6, infra).

In addition, gene expression is rapid 30 because it does not require the entry of the DNA into the cell nuclei for the initiation of transcription. The cytoplasmic location of its site of action also provides for a safety feature which reduces the possibility of its integration into the host cell 35 genome at a site adjacent to and thereby activating a dormant gene such as an oncogene. Furthermore, unlike retroviral vectors, even if DNA integration occurred,

the exogenous promoter used would not be activated by eukaryotic transcription proteins. The only RNAP capable of activating the promoter sequence to initiate transcription is synthesized in the cell 5 cytoplasm, and would not be able to gain access to the nucleus for binding to its cognate promoter because it lacks any nuclear translocation signal.

This expression system encompasses a variety of applications. For example, it is especially 10 suitable for expressing genes in a specific cell type where appropriate promoters for that cell type are unavailable. It is also useful in vitro for the transient expression of a newly cloned gene for confirmation of the identity of its encoded gene. 15 product as well as the rapid production of high amounts of proteins for use in immunization of animals for the generation of specific antibodies. In this regard, the present invention may be used to introduce exogenous genes into cells and tissues in vivo for 20 transient expression of the gene product to elicit an antigen-specific host immune response. The short-term nature of gene expression may be ideal for its use as a vehicle for in vivo immunization of a host to an antigen.

25 A major impediment in the current attempts of gene therapy is the integration of foreign genes in non-dividing eukaryotic host cells. The ability of the present system to permit gene expression in the cell cytoplasm circumvents this potential problem, as 30 gene expression can be achieved in quiescent cells. However, as the DNA construct is gradually degraded in the cytoplasm or diluted in number when cells divide over time, the present invention is also self-limiting. The self-limiting nature of this expression 35 system is particularly suitable for use in settings

where gene expression is desirable only temporarily or transiently. For example, the present invention may be used to target lymphokines genes to tumor sites in vivo for the in situ activation of cytotoxic 5 lymphocytes to mediate tumor cytolysis. Since the expression of the exogenous genes is transient, the uptake of DNA by cells not lysed in the process would not eventually induce permanent and sustained synthesis of lymphokines leading to potentially 10 deleterious consequences, such as autoimmunity.

The present invention encompasses an expression system used as a single unit or as a dual system. A single-unit system includes the RNAP gene and a gene of interest within the same DNA construct 15 prebound to RNAP. A dual system includes an autogene containing a RNAP prebound by RNAP and a separate plasmid containing a gene of interest driven by the same cognate RNAP promoter also prebound by RNAP. In many circumstances, it is most convenient to introduce 20 a single DNA construct into host cells. However, the two plasmid systems are particularly useful in settings where it is desirable to adjust the ratio of the expressed RNAP and the gene product of interest by providing different ratios of the two plasmids. In 25 this way, the system prevents uncontrolled expression of one protein over the other, especially if the RNAP is synthesized at a much faster rate than the gene of interest.

The expression system may be complexed with 30 RNAP and suspended in saline, PBS, serum free media or any aqueous solution suitable for in vivo administration. The solution may be injected into an animal, including a human, intravenously, intramuscularly, intracranially, subcutaneously or 35 directly into a tissue or organ. The DNA-RNAP complex

may be injected alone, encapsulated by liposomes or linked to any carrier molecules capable of translocating the complex across the plasma membrane. For in vivo administration, the expression system may 5 be injected in the dose range of 0.01-100 mg/kg. If the gene product of interest is desired to be secreted into the bloodstream, the complex may be injected intravenously to cause endothelial cell uptake and release of the gene product directly into the 10 circulation.

6. EXAMPLE: T7 RNA POLYMERASE PREBOUND TO
DNA DIRECTS GENE EXPRESSION IN
CULTURED CELL LINES

15 6.1. MATERIALS AND METHODS

6.1.1. ENZYMES

Bacteriophage T7 RNAP (50U/ul), restriction endonucleases, and the Klenow fragment were purchased from New England BioLabs.

20

6.1.2. CELLS AND PLASMIDS

Mouse L and human HepG2 cells were maintained in DMEM supplemented with 10% Nu serum or 10% calf serum (growth media), respectively. E. coli 25 DH5 α was from Bethesda Research Laboratories. E. coli HMS174 with pLysE (Studier et al., 1990, Methods Enzymol. 185:60-89; Studier, 1991, J. Mol. Biol. 219:37-44); plasmid pAR1173 (Davanloo et al., 1984, Proc. Natl. Acad. Sci. USA 81:2035-2039), which 30 contains a gene coding for T7 RNAP, and pATUO-1 (Dubendorff and Studier, 1991, J. Mol. Biol. 219:61-68), which contains a Lac I gene, were provided by Dr. Studier. Plasmid pTM-I (Moss et al., 1990, Nature 348:91-92) was from Dr. Moss' laboratory. 35 Plasmid phGH (Martíal et al., 1979, Science

205:602-607) was from Dr. Goodman's laboratory. Plasmid pBLCAT (Luckow and Schutz, 1987, Nucleic Acids Res. 15:5490), which contains a CAT gene, was obtained from Dr. T. Coleman; and pMThGH, which contains hGH 5 cDNA (Martial et al., 1979, Science 205:602-607) driven by a mouse metallothionein I promoter (Brinster et al., 1981, Cell 27:223-231), was constructed in our laboratory. The bacteria were grown in LB medium with appropriate antibiotics.

10

6.1.3. CONSTRUCTION OF PLASMIDS

pTM-I, a cytoplasmic expression vector which contains a T7 promoter connected to at its 3' end an EMC capping independent sequence (Moss et al., 1990, 15 Nature 348:91-92), was linearized with restriction enzyme Bam HI. A 2.6 kbp Bam HI fragment of pAR1173 containing a T7 gene was inserted into pTM-I vector by ligation. The ligation products were subsequently transformed into E. coli strain DH5 α . However, 100% 20 of the recombinant clones were found to have T7 gene inserted in an orientation opposite to that of the promoter's (which formed a T7T7 construct, not T7T7), suggesting that T7T7 construct was lethal to host bacteria (Studier, 1991, J. Mol. Biol. 219:37-44; 25 Dubendorf and Studier, 1991, J. Mol. Biol. 219:61-68; Dubendorff and Studier, 1991, J. Mol. Biol. 219:45-59). Several steps were subsequently taken to reduce the toxicity to bacterial hosts resulted from the expression of the T7 gene, while maintaining the 30 ability of the construct to be expressed in eukaryotic cells.

First, a 1.6 kbp EagI/EcoNI Lac I gene which encodes a repressor was isolated from pAUTO-1 and inserted into ClaI and XbaI sites of pTM-I by a 35 blunt-end ligation (FIG. 2). In addition, an operator

sequence, GGA ATT GTG AGC GGA TAA CAA TTCC (25mer), which provides the binding site for Lac I repressor, was inserted immediately 3' to the T7 promoter sequence (as shown in pTLO-I in FIG. 2) where the 5 operator was shown to have the maximal suppression of T7 promoter activity (Dubendorff and Studier, 1991, J. Mol. Biol. 219:45-59). As a second step to further reduce the cytotoxicity, the Shine-Dalgarno sequence (S-D box) of the T7 gene was removed from the gene to 10 reduce unwanted translation of T7 mRNA which was generated as a result of a leaky Lac I suppression (FIG. 2). Oligonucleotide CCC GAT TTA CTA ACT CCA TGG ACA CGA TTA ACA TCG CTA AG (41mer) was used as a primer for deletion of S-D box sequence using a 15 phagemid mutagenesis method (Kunkel, 1985, Proc. Natl. Aca. Sci. USA 82:488-492; Kunkel et al., 1987, Methods Enzymol. 154:367-382). In addition, an NcoI site (CCATGG, as underlined in the oligonucleotide) was added to the T7 gene. A 2.6 kbp NcoI/BamHI modified 20 T7 fragment was inserted into the gap between the NcoI and BamHI sites of pTM-I in such a manner that the optimized translation of the T7 gene would be initiated from the ATG in the NcoI site (Brinster et al., 1981, Cell 27:223-231) (FIG. 2). However, this 25 modification altered the second amino acid residue of T7 RNAP from an Asn (AAC) to an Asp (GAC). Finally, the constructed pT7T7 was transformed into and prepared from HMS174 cells which contained a plasmid pLysE encoding a T7 lysozyme, a T7 RNAP inhibitor. 30 All the mutations described herein were confirmed by DNA sequencing (Sanger et al., 1977, Proc. Natl. Acad. Sci. USA 76:4350-4354).

Construction of pT7hGH was simpler than that of pT7T7 since hGH is not toxic to the bacterial 35 hosts. pTM-I was linearized with SmaI in the

polycloning site. A 0.9 kbp Hind III fragment of an hGH cDNA was isolated from plasmid phGH, 5' protruding ends of the fragment were filled by Klenow fragment and dNTPs, and was inserted into pTM-I by a blunt-end 5 ligation. With a deletion mutation, the reading frame of the hGH sequence was adjusted in a similar manner as for PT7T7 so that the ATG in NcoI site serves as the first codon for the hGH cDNA (FIG. 3A).

Construction of PT7CAT was carried out in a 10 similar fashion as that of pT7hGH. A 1.7 kbp DNA fragment containing CAT gene was isolated from pBLCAT3, blunt ended with Klenow fragment and dNTPs, and inserted into SmaI site of pTM-I by ligation. The relative position of ATG codon of CAT gene to the T7 15 promoters was also adjusted to the optimal position by a deletion mutation (FIG. 3B). The adjustments for both hGH and CAT genes did not result in mutations in their amino acid sequences.

Following construction of pT7T7 and pT7hGH, 20 a single plasmid, pT7T7/T7hGH, was constructed from these sequences. As shown in FIG. 4, pT7T7 was linearized at EagI site, the EagI site was subsequently filled by Klenow fragment. A 2.1 kbp ClaI/EagI T7hGH fragment was isolated from pT7hGH, the 25 single stranded cohesive ends were also filled in by Klenow fragment. pT7T7/T7hGH was made by ligating these two sequences together followed by transformation using HMS174 pLysE cells. Large scale preparation of pT7T7/T7hGH, and other plasmid DNA used 30 in these experiments, were made using Qiagen columns (Qiagen).

6.1.4. ADDITION OF POLY(dT) TAILS

The T7 system described herein was designed as a cytoplasmic expression system and therefore no natural poly(A) tails would be added to the hGH and 5 CAT mRNA generated by this system. Poly dT tails of 40 base pairs long were added to the 3' ends of both hGH and CAT genes in order to increase the stability and possibly the translation efficiency of the hGH and CAT mRNA.

10

6.1.5. TRANSFECTION

Mouse L and human HepG2 cells were grown in growth media in 6-well cell culture dishes to 80% confluence. Plasmid DNA was diluted with water and 15 the T7 buffer (New England Biolabs) to various concentrations in a total volume of 75 μ l in sterile polystyrene tubes. Alternatively, the T7 buffer may be replaced by saline, PBS or DMEM. Five μ l of T7 RNAP (50U/ μ l) was added to DNA solution and the 20 mixture was incubated at room temperature for 10 min, followed by addition of 20 μ l (1 μ g/ μ l) of lipofectin (Felgner et al., 1987, Proc. Natl. Acad. Sci. USA 84:7413-7417). After gentle mixing, the lipofectin-DNA-enzyme solution was incubated at room 25 temperature for another 5 min. Meanwhile, growth media were removed from L cells, the cells were washed twice with DMEM. One and half ml of DMEM was added to each well, followed by addition of 100 μ l of lipofectin-DNA-enzyme solution. After gentle mixing, 30 the dishes were incubated at 37°C in a cell culture incubator. Following 4 hours of incubation, the media were removed, 2ml of growth media were added to each well, and the dishes were incubated at 37°C with 5% CO_2 . Growth media from the transfected cells were 35 collected every 8 hours for the first 24 hours and

- 30 -

then every 24 hours and the cells were replenished with fresh growth media. Cells which were transfected with pT7T7/T7hGH alone, pT7hGH + T7 RNAP, or pMThGH were used as expression controls. pMThGH, a 5 conventional expression vector, also served as a reference for comparison of expression profiles. Transfection of pMThGH was performed using a standard lipofection protocol without adding T7 RNAP. In a similar manner, pT7CAT and pT7CAT + T7 RNAP were used 10 as controls for CAT assays, respectively. In DEAE-dextran transfections, DNA-T7 RNAP solution was prepared identically as described for lipofection. After adding DEAE-dextran containing DMEM to DNA-T7 15 enzyme solution, cells were transfected under the conditions as previously described (Chen et al. (1992) Mol. Endocrinol. 6:598-606).

6.1.6. RADIOIMMUNOASSAY AND CAT ASSAY

hGH RIA was performed using a commercially 20 available RIA kit (Hybritech, San Diego). pT7T7, pT7CAT, and T7 RNAP were coincubated and transiently transfected into either L or hepG2 cells by the same lipofection protocol as described. Twenty four hours after the transfection, the cells from each well of a 25 6-well dish were individually harvested in 1 ml PBS, centrifuged and resuspended in 100 μ l of 100 mM Tris (pH 7.8). Frozen and thawed three times, the cell debris was removed by centrifugation. The supernatant was incubated at 65°C for 10 min, followed by 30 centrifugation to remove protein precipitates. The resulting supernatant was assayed for CAT activity as described (Neumann et al., 1987, Biotechniques 5:444-447).

6.1.7. NORTHERN BLOT ANALYSIS

Twenty four or 48 hours after the transfection, transfected cells were lysed by 1 ml of RNAzol (Cinna/Biotecx Laboratories) immediately after 5 PBS wash. Total RNA was isolated as described (Chomczynski and Sacchi, 1987, Anal. Biochem. 162:156-159). Twenty μ g of total RNA from each cell sample was subjected to 1% formaldehyde gel electrophoresis. Afterwards, resolved RNA was 10 transferred from the gel to a nylon-based membrane (Gene Screen Plus from NEN), hybridized to a 32 P-labeled, 2.6 kbp T7 fragment and subsequently visualized by autoradiography (Rave et al., 1979, Nucleic Acids Res. 6:3559-3567). After being stripped 15 off the T7 probe, the same membrane was rehybridized to a 0.9 kbp hGH probe. Actin mRNA served as reference controls to standardize sample signal intensities.

20

6.2. RESULTS6.2.1. CONSTRUCTION OF PLASMIDS

Figures 2, 3A, 3B and 4 schematically demonstrate the construction of the plasmids, pT7T7, pT7hGH, pT7CAT, and PT7T7/T7hGH. The pT7T7 and 25 pT7T7/T7hGH plasmids were grown in the presence of the pLysE plasmid to reduce the toxicity of the T7 enzymes to E. coli. Therefore, these plasmid preparations were always contaminated with some pLysE plasmid (< 10%). Since pT7CAT and pT7hGH were not toxic to their 30 bacterial hosts, their replication did not require pLysE.

Efforts were made to construct T7T7 sequence in order to reduce the cytotoxicity to such levels as to allow sufficient quantities of the plasmids to be 35 prepared from bacterial cells for eukaryotic

expression studies. Most of the genetic manipulations were similar to those used in construction of T7 vectors for the expression of cloned genes in bacteria (Studier, 1991, J. Mol. Biol. 219:37-44; Dubendorff et al., 1991, J. Mol. Biol. 219:61-68; Dubendorff et al., 1991, J. Mol. Biol. 219:45-59). However, the goal of making expression systems functioning in eukaryotic cells, not in bacterial cells, permitted more genetic modifications such as the removal of S-D box from the 10 T7 gene, to further minimize its toxicity to E. coli host cells (FIG. 2). These manipulations are advantageous with respect to increasing plasmid yields and reducing the risk of DNA mutations resulted from the selective pressure exerted upon the host cells by 15 the toxic effects of the expressed T7 RNAP.

6.2.2. COMPARISON OF LIPOFECTIN AND DEAE-DEXTRAN TRANSFECTIONS

In order to determine which DNA delivery methods worked more efficiently for the T7 system, both lipofectin or DEAE-dextran transfection methods were compared on the same 6-well cell culture plates. Table I shows the relative transfection efficiencies of lipofection and DEAE-dextran methods on the T7 expression systems. CAT assays, compared to hGH assays, resulted in smaller standard deviations largely due to the higher sensitivity of the assays while hGH assays could not as accurately detect low levels of hGH expression from DEAE-dextran transfected 20 cells. As indicated by Table I, lipofection was approximately 8-10 times more efficient than 25 DEAE-dextran method for the T7 expression system.

The key for the success of the expression of the T7 systems in eukaryotic cells is to keep the T7 30 RNAP tightly bound in undegraded forms to the DNA during the process of transfection. It is possible

that liposomes better protected the DNA-T7 enzyme complex and/or better facilitated the complex to reach the cytoplasm of the cells.

5

TABLE I

Comparison of relative transfection efficiencies.

10	Transfection Method	Relative hGH activity	Relative CAT Activity
	Lipofection	100	100
	DEAE-dextran	11.5±4.6	14.2±3.9

15

6.2.3. TRANSIENT EXPRESSION OF THE FUNCTIONAL/REPORTER GENES

6.2.3.1. hGH EXPRESSION

20 The expression of pT7T7+pT7hGH (dual plasmid) or pT7T7/T7hGH (single plasmid) in L cells as compared to pT7hGH and pMThGH is shown in FIG. 5. Cell culture supernatants from transiently transfected L cells were collected and assayed for hGH every 8 hours for the first 24 hours and then every 24 hours 25 for 5 days. By day 7, the cells were severely overgrown in the well and started to die.

30 Detectable levels of hGH were found as early as 8 hours after transfection in cell culture fluids collected from cells transfected either by pT7T7 + pT7hGH + T7 RNAP or pT7T7/T7hGH + T7 RNAP. The hGH in the culture medium of transfected L cells was biologically active in stimulating rat Nb2 cells. However, expression of pMThGH (which served as 35 positive control as well as a reference for expression profiles in this study) could not be detected until 24

hours after the transfection. The expression of hGH by pMTbGH was not higher than that of the T7 system until 72 hours or later. For cell samples transfected by pT7hGH plus T7 RNAP (negative control I), only the 5 culture medium collected 8 hours after the transfection demonstrated low levels of hGH (FIG. 5). The expression disappeared after longer incubation periods. This indicates that it was the post-transfection expression of the T7 gene that 10 maintained expression of the hGH gene. No hGH could be detected at any time in the culture fluids collected from the cells transfected by pT7T7/T7hGH alone without prebound T7 RNAP. The expression of the T7 system reached its peak (~3 ng/ml) approximately 24 15 to 48 hours after the transfection whereas expression of pMThGH reached its peak (~7ng/ml) at about 96 to 120 hours. Therefore, the kinetics of the hGH expression by the T7 system seemed to be quite different from that of a traditional expression 20 vector.

The onset of expression of hGH using the T7 system was more rapid than that of a conventional mammalian promoter-containing plasmid pMThGH. This result was consistent with studies using a T7CAT 25 vector coupled with recombinant vaccinia viruses, which showed that 48 hours after the transfection the cells transfected by the T7 system produced several hundred-fold higher CAT activities than those expressed either RSVCAT or SV40CAT (Fuerst et al., 30 1986, Proc. Natl. Acad. Sci. USA 83:8122-8126). The rapid expression and different kinetics exhibited by the T7 system indicate that an expression mechanism, which differs from the one used by pMTbGH, was employed by the T7 system. It is likely that the 35 plasmid-bound T7 RNAP initiated transcription

immediately after the plasmids were taken into the cytoplasm of the cells, followed by rapid protein synthesis and secretion which resulted in the shift the expression curves to the left. In contrast, 5 pMTbGH had to reach the nuclei of the transfected cells for the hGH expression which may explain the long delay of the hGH expression by pMThGH. This result is consistent with those of other studies which showed that reporter gene was actively transcribed by 10 T7 RNAP in the cytoplasm of cells (Fuerst et al., 1986, Proc. Natl. Acad. Sci. USA 83:8122-8126; Elroy-Stein et al., 1990, Proc. Natl. Acad. Sci. USA 87:6743-6747). It is also consistent with the cytoplasmic expression nature of the T7 parental 15 plasmid pTM-I (Moss et al., 1990, Nature 348:91-92). Because of the rapid and efficient expression in the early hours post transfection compared to other conventional plasmid systems, this T7 expression system may prove to be useful to express cDNA in those 20 circumstances in which rapid assays are desirable.

6.2.3.2. CAT EXPRESSION

Figure 6 demonstrates the results of CAT assays. Cells were lysed and assayed 48 hours after 25 the transfection. CAT activities in pT7T7 + pT7CAT + T7 RNAP transfected cells were over 50 times more than that of the PT7CAT + T7 RNAP transfected samples (negative control type I) and were more than 100 times that of pT7CAT transfected samples (negative control 30 type II). On the basis of absolute quantity, it turned out that the intracellular CAT activity corresponded to approximately 2-3 ng CAT protein/well (or per 10^6 cells), in the same range of magnitude as extracellular hGH levels expressed by either 35 PT7T7/T7hGH+T7 RNAP or pT7T7+pT7hGH+T7 RNAP.

6.2.4 NORTHERN ANALYSIS OF T7 mRNA

In order to prove that the sustained expression of the functional/reporter genes in the T7 system was indeed due to the sustained expression of the T7 genes in the same system, total RNA from the transfected cells was isolated and analyzed by Northern blots as shown in FIG. 7A and 7B. Both T7 RNAP and hGH mRNA were found in the cells transfected by pT7T7/T7hGH+T7 RNAP, but not in the cells transfected either by pT7T7/T7hGH alone or by pT7hGH+T7 RNAP (FIG. 7A and 7B). The position of the major bands on the blots corresponded to the anticipated sizes of the T7 RNAP and hGH mRNA. Also, the levels of T7 and hGH mRNA were found to correlate with each other, i.e., the higher the T7 mRNA level (24 hr), the higher the hGH mRNA level, and vice versa (48 hr samples).

20

6.2.5. EXPRESSION OF hGH AND CAT IN BOTH L AND HepG2 CELLS

Expression of hGH and CAT by mouse L and human HepG2 cells transfected by pT7T7+pT7hGH or pT7T7+pT7CAT is summarized in Table II. Because the two cell types grew in very different modes (L cells grew in a single uniform layer whereas HepG2 cells grew in clusters), hGH and CAT levels per well were found to be quite different for the two cell types. However, when the expression levels were adjusted on the basis of each mg of cell proteins, it was found that L cells and HepG2 cells expressed both genes, particularly the CAT gene, with comparable efficiencies (Table II).

TABLE II
Relative expression efficiencies
by different cell lines.

5	Relative Cell lines	Relative hGH activity	Relative CAT Activity
	Mouse L cells	100	100
10	Human HepG2 cells	268 \pm 51	177 \pm 25

7. EXAMPLE: T7 RNA POLYMERASE PREBOUND
TO DNA DIRECTS GENE EXPRESSION
IN IN VIVO TISSUES

15

7.1. MATERIALS AND METHODS

7.1.1. CONSTRUCTION OF PLASMID

In order to construct a pT7T7/T7Luc plasmid, a pT7Luc plasmid was first constructed by insertion of a luciferase cDNA (de Wet et al., 1987, Mol. Cell.

20 Biol. 7:725-737) into pTM-I, followed by cleavage of ClaI/EagI T7-luciferase fragment from pT7Luc, and insertion of the blunt-ended T7-luciferase fragment into blunt-ended EagI site of pT7T7.

25

7.1.2. IN VIVO ADMINISTRATION

Varying concentrations of plasmids pT7T7/T7Luc and pT7T7/T7hGH were complexed with T7 RNAP in a solution with or without lipofectin and injected directly into mouse tail, liver, leg muscles,

30 cerebrospinal fluids or intravenously. Injection was performed by injecting 70-100 μ l of DNA-T7 RNAP in saline, PBS or DMEM into animal tissues using 25-gauged needles. After a certain time period, gene expression was assessed by sacrificing the animals and

35 a portion of specific tissues was removed. In the

case of luciferase expression, the tissue was homogenized in 200-400 μ l of luciferase lysis buffer, and 10 μ l of the tissue extract was assayed for luciferase activities using a commercially available 5 kit from Promega. In the case of growth hormone expression, cerebrospinal fluids, brain tissues and sera were assayed for hGH activity by commercially available radioimmunoassay kits.

10

7.2. RESULTS

In order to facilitate the detection of gene expression *in vivo*, the luciferase gene was chosen and inserted into the expression system of the invention as pT7T7/T7Luc. When the plasmid DNA was complexed 15 with T7 RNAP *in vitro* and injected with Lipofectin into the connective tissues of the tail of mice subcutaneously, a high level of luciferase activity was detected in homogenized tail tissues 24 hours after injection (FIG. 8). This response was dose- 20 dependent, and luciferase gene expression was detectable for at least one week. Control plasmids did not produce detectable activities. When the same material was injected into the tail vein of mice intravenously, and the tail vein was dissected from 25 the rest of the tail tissues about 24 hours later, the luciferase activity detected in these cells was over a million times more than that in the tail tissues, as compared by tissue weight. This indicates that intravenous injection of DNA-RNAP complex may be taken 30 up immediately by endothelial cells leading to rapid expression of the gene product, especially near the site of injection.

Similarly, when pT7T7/T7Luc with prebound T7 RNAP was injected into the mouse leg muscles 35 intramuscularly, into mouse livers intrahepatically or

into the cerebrospinal fluids of 20 day-old mice intracranially, significant luciferase activity was observed in muscle tissues (FIG. 9), in mouse livers (FIG. 10) and in brain tissues (FIG. 11).

5 The pT7T7/T7hGH plasmid encoding secreted human growth hormone was also tested in vivo. The DNA was complexed with T7 RNAP and liposome, and injected into the cerebrospinal fluids of 20 day-old mice. Growth hormone activity was detected in both 10 cerebrospinal fluids and brain tissues from the injected animals 20 hours after injection (FIG. 12).

In addition, when 100 µg of pT7T7/T7hGH plasmid complexed with T7 RNAP in the absence of 15 lipofectin was injected into the connective tissues of the tail of mice subcutaneously, 200-300 pg/ml of human growth hormone was detected in the peripheral blood the following day.

The present invention is not to be limited 20 in scope by the exemplified embodiments which are intended as illustrations of single aspects of the invention. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the 25 art from the foregoing description and accompanying drawings. Such modifications are intended to fall within the scope of the appended claims.

All publications cited herein are incorporated by reference in their entirety.

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into the cerebrospinal fluids of 20 day-old mice intracranially, significant luciferase activity was observed in muscle tissues (FIG. 9), in mouse livers (FIG. 10) and in brain tissues (FIG. 11).

5 The pT7T7/T7hGH plasmid encoding secreted human growth hormone was also tested in vivo. The DNA was complexed with T7 RNAP and liposome, and injected into the cerebrospinal fluids of 20 day-old mice. Growth hormone activity was detected in both 10 cerebrospinal fluids and brain tissues from the injected animals 20 hours after injection (FIG. 12).

In addition, when 100 μ g of pT7T7/T7hGH plasmid complexed with T7 RNAP in the absence of 15 lipofectin was injected into the connective tissues of the tail of mice subcutaneously, 200-300 pg/ml of human growth hormone was detected in the peripheral blood the following day.

The present invention is not to be limited 20 in scope by the exemplified embodiments which are intended as illustrations of single aspects of the invention. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the 25 art from the foregoing description and accompanying drawings. Such modifications are intended to fall within the scope of the appended claims.

All publications cited herein are incorporated by reference in their entirety.

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15. The expression system of Claim 13 in which the gene of interest encodes chloramphenicol acetyltransferase.

5 16. The expression system of Claim 13 in which the gene of interest encodes luciferase.

10 17. The expression system of Claim 12 in which the gene of interest encodes an anti-sense RNA.

18. The expression system of Claim 12 in which the gene of interest encodes a ribozyme molecule.

15 19. A method for expressing an exogenous gene in eukaryotic cells comprising co-introducing into host cells:

20 (a) a recombinant DNA molecule which comprises a coding sequence for an RNA polymerase or a functional derivative thereof, operably linked to its cognate promoter, which DNA molecule is prebound to the RNA polymerase capable of initiating transcription from its cognate promoter; and

25 (b) a second recombinant DNA molecule which comprises a gene of interest or a functional derivative thereof, operably linked to a second copy of the cognate promoter of (a).

30 20. The method of Claim 19 in which the second recombinant DNA molecule is prebound to the RNA

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polymerase capable of initiating transcription from its cognate promoter.

21. The method of Claim 20 in which the RNA 5 polymerase coding sequence is derived from bacteriophage T7.

22. The method of Claim 21 in which the gene of interest encodes a protein.

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23. The method of Claim 22 in which the gene of interest encodes human growth hormone.

24. The method of Claim 22 in which the 15 gene of interest encodes human chloramphenicol acetyltransferase.

25. The method of Claim 22 in which the gene of interest encodes luciferase.

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26. The method of Claim 21 in which the gene of interest encodes an anti-sense RNA.

27. The method of Claim 21 in which the 25 gene of interest encodes a ribozyme molecule.

28. A method for expressing an exogenous gene in eukaryotic cells comprising introducing into host cells a recombinant DNA molecule which comprises 30 a coding sequence for an RNA polymerase or a functional derivative thereof, operably linked to its cognate promoter, and a gene of interest or a functional derivative thereof, operably linked to a second copy of the cognate promoter, which DNA

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molecule is prebound to the RNA polymerase capable of initiating transcription from its cognate promoter.

29. The method of Claim 28 in which the RNA polymerase coding sequence is derived from 5 bacteriophage T7.

30. The method of Claim 29 in which the gene of interest encodes a protein.

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31. The method of Claim 30 in which the gene of interest encodes human growth hormone.

32. The method of Claim 30 in which the 15 gene of interest encodes chloramphenicol acetyltransferase.

33. The method of Claim 30 in which the gene of interest encodes luciferase.

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34. The method of Claim 29 in which the gene of interest encodes an anti-sense RNA.

35. The method of Claim 29 in which the 25 gene of interest encodes a ribozyme molecule.

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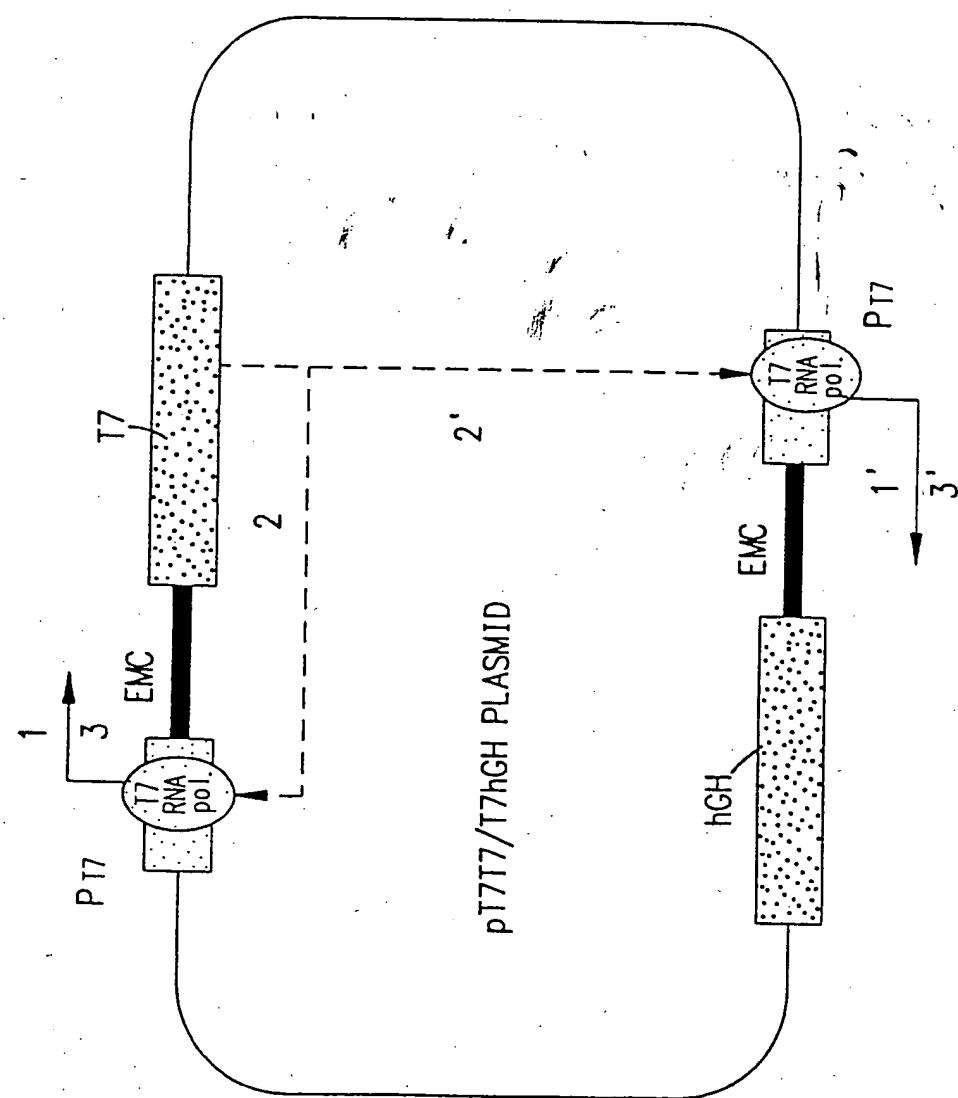


FIG. 1

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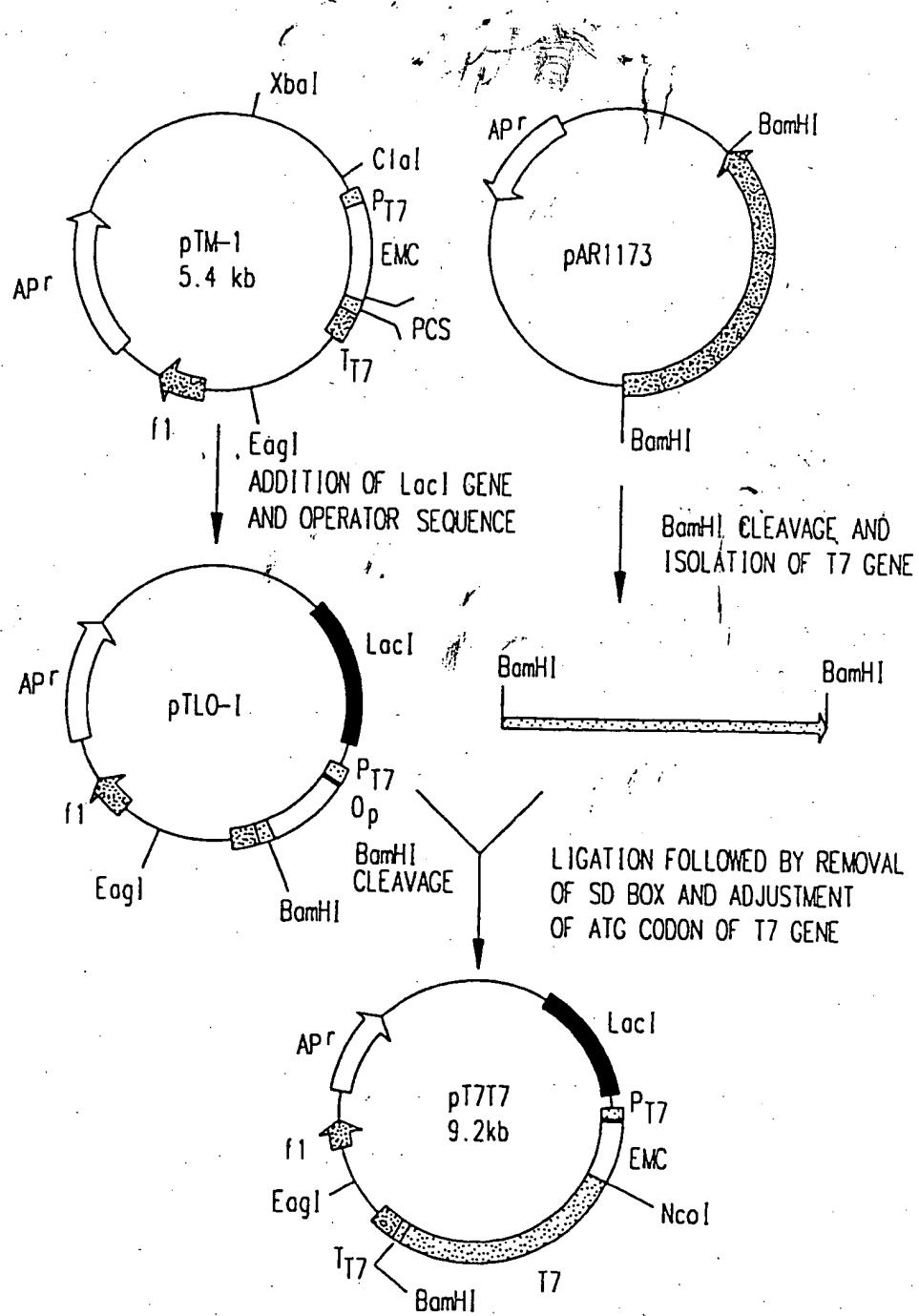


FIG.2

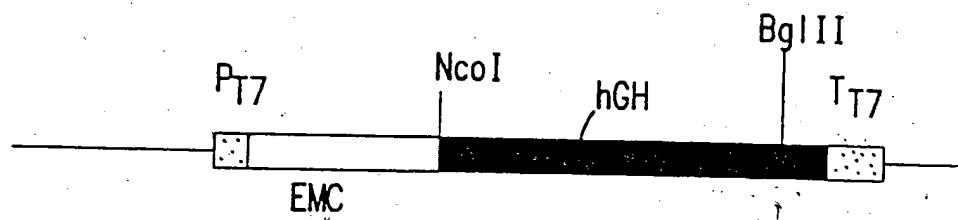


FIG.3A

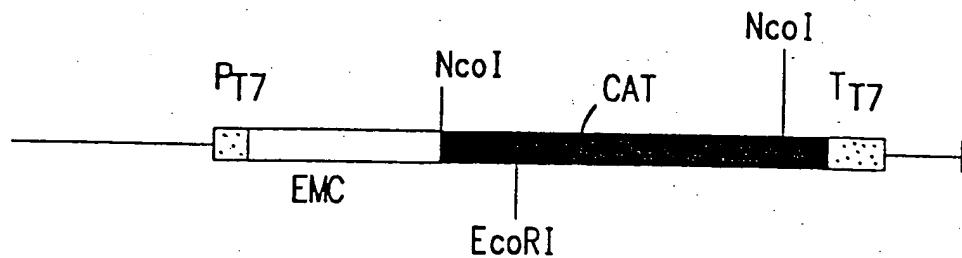


FIG.3B

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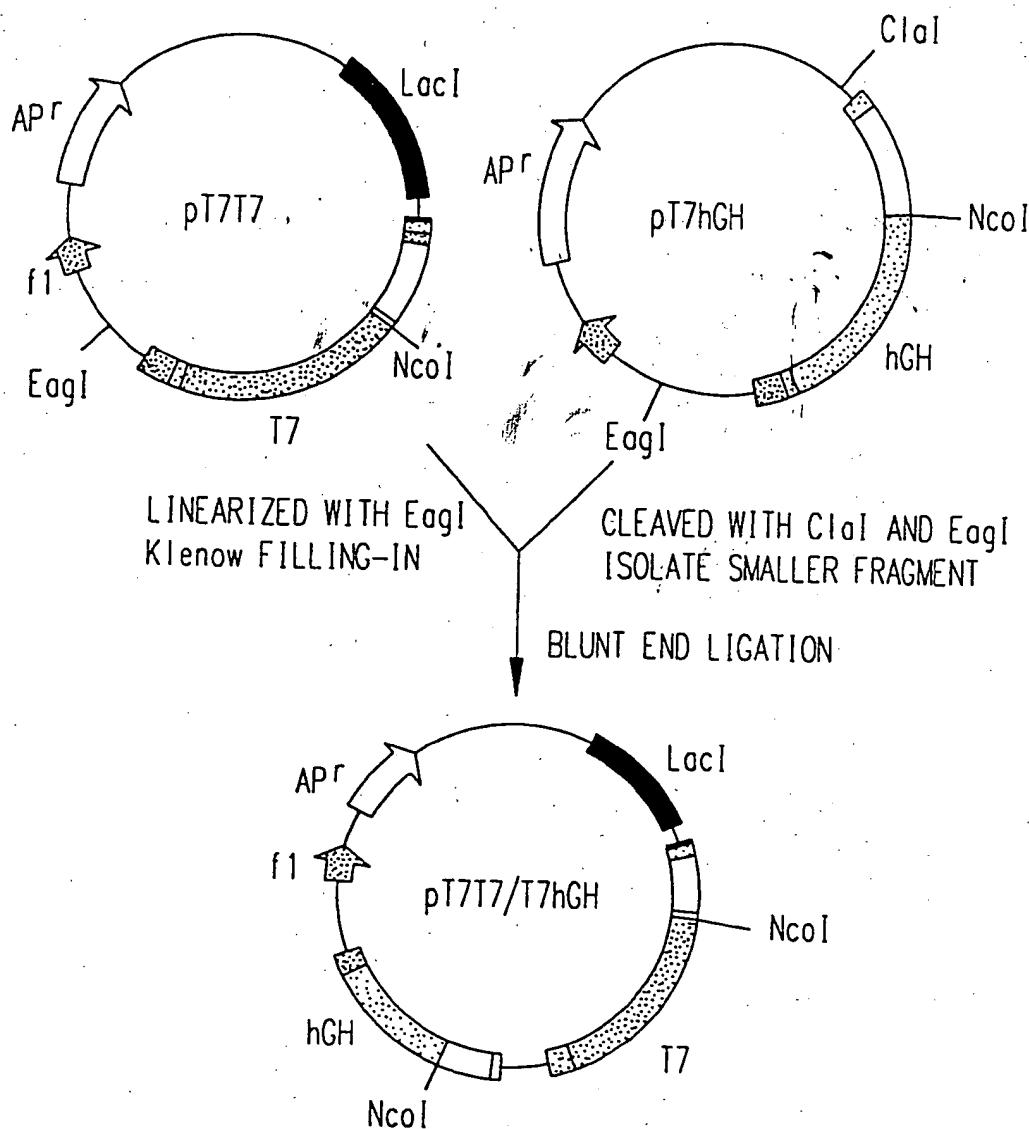


FIG.4

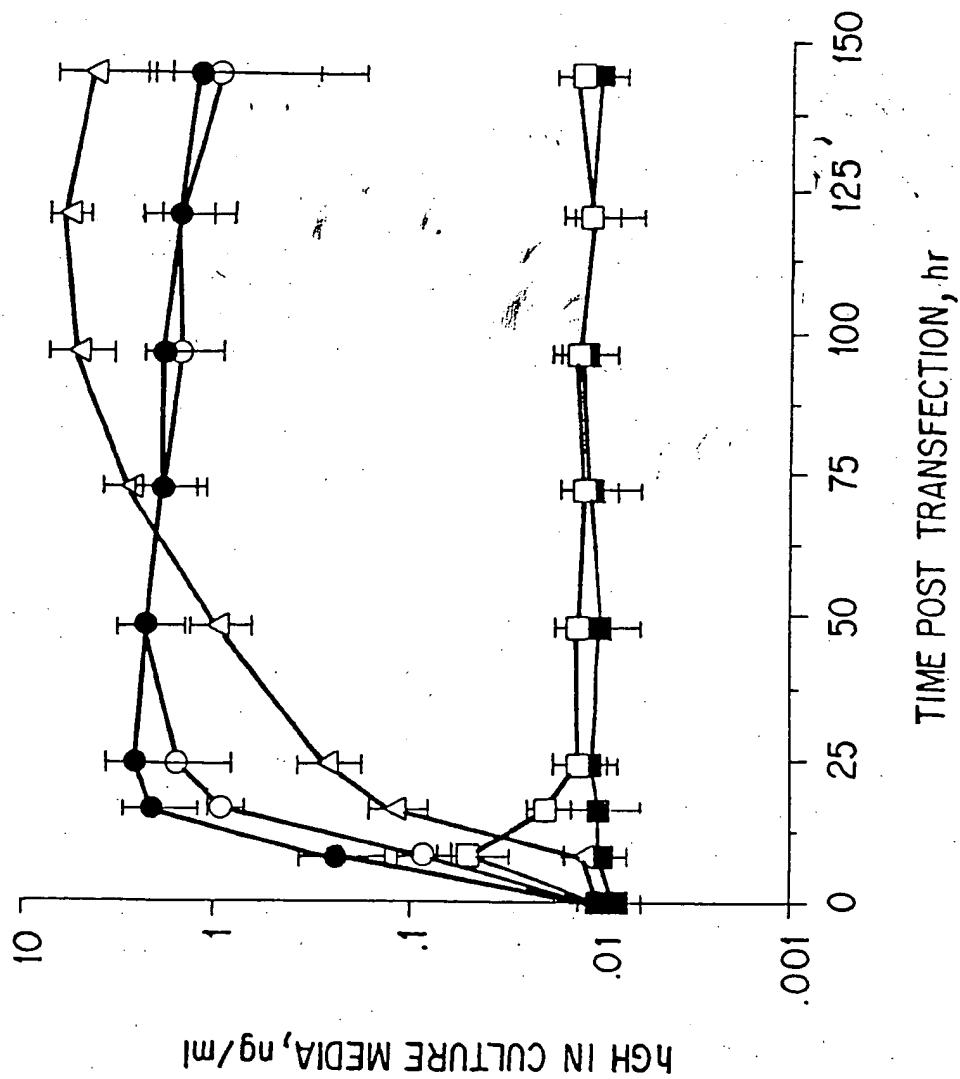


FIG. 5

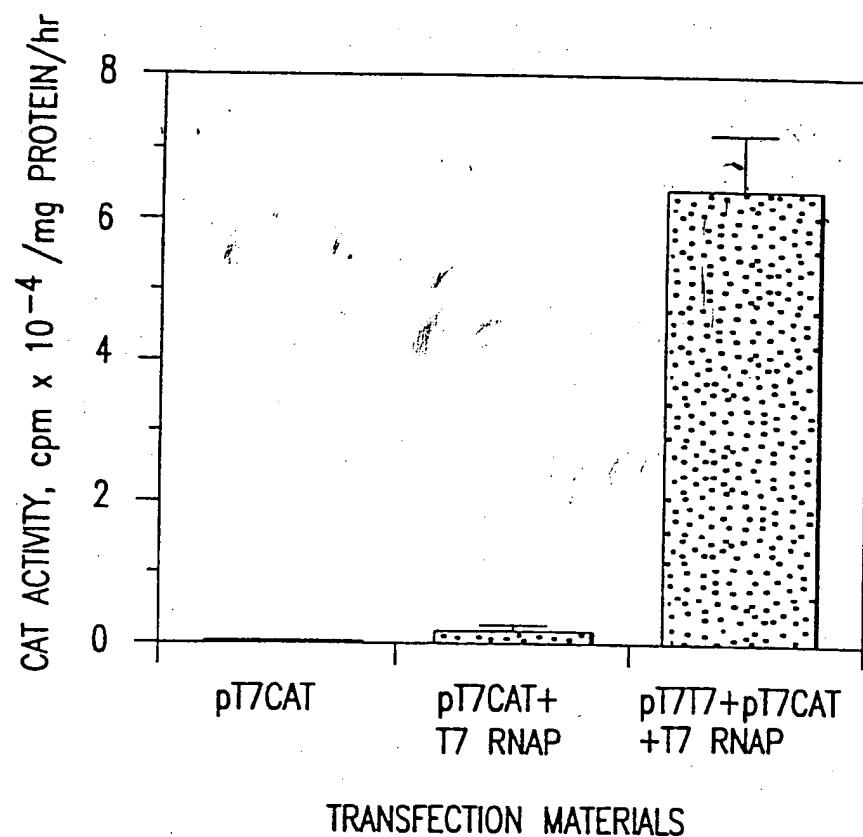


FIG.6

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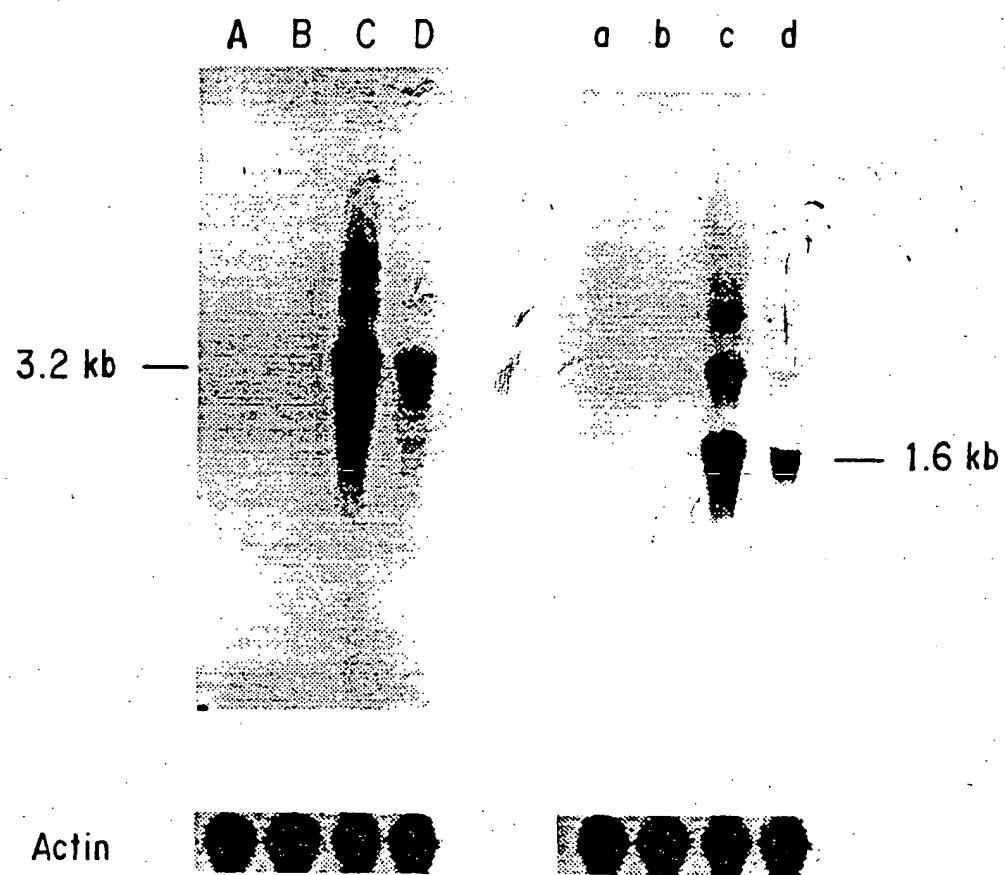


FIG. 7A

FIG. 7B

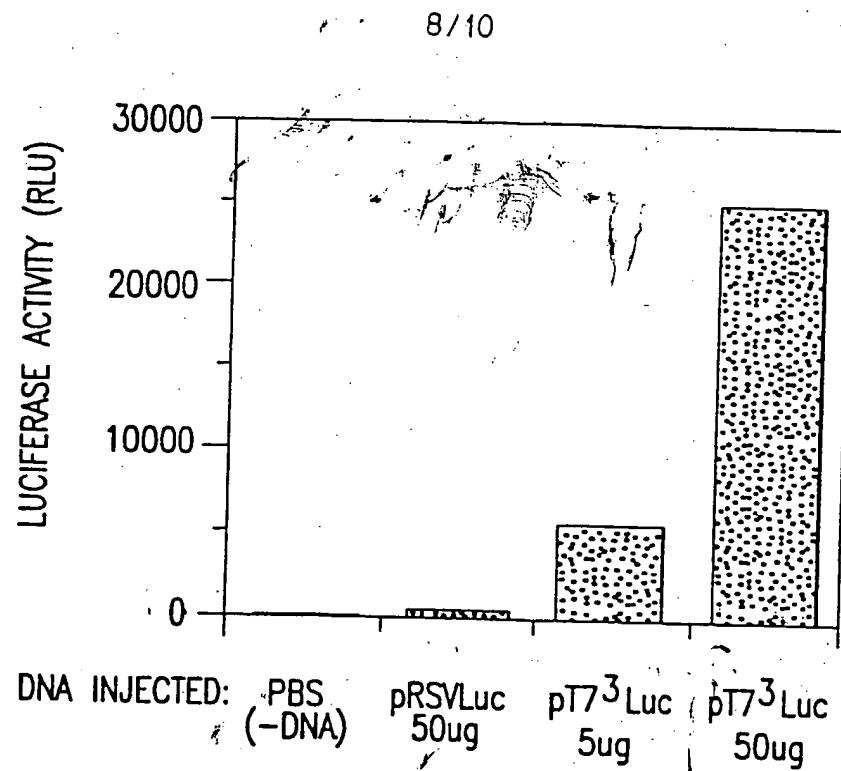


FIG.8

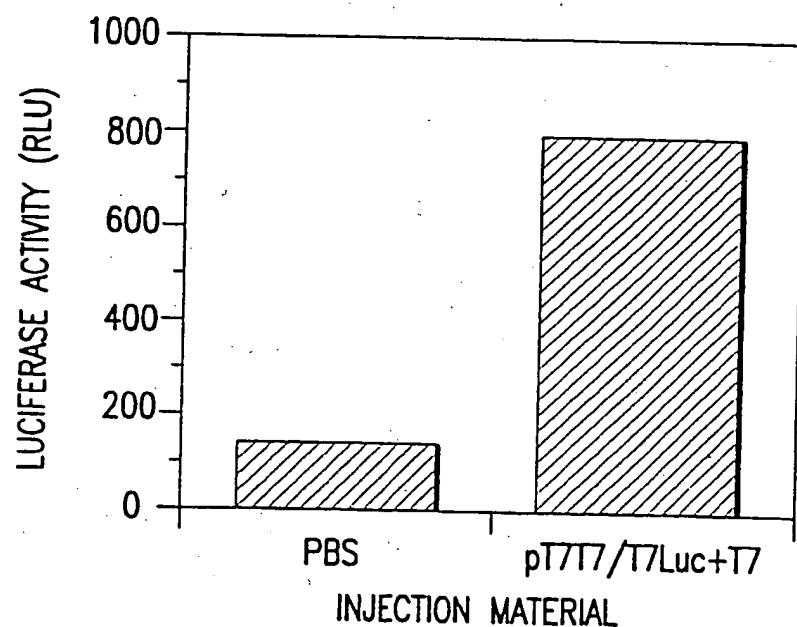


FIG.9

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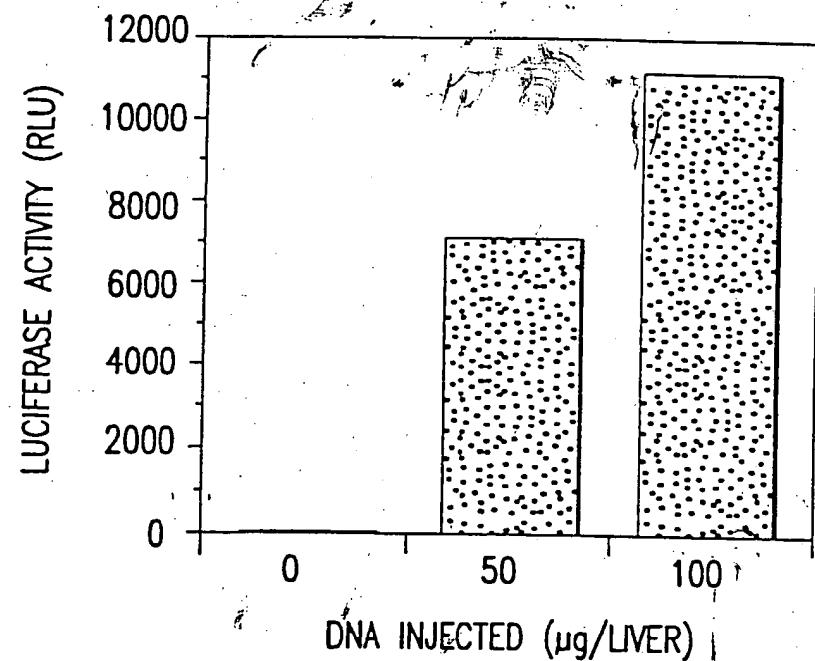
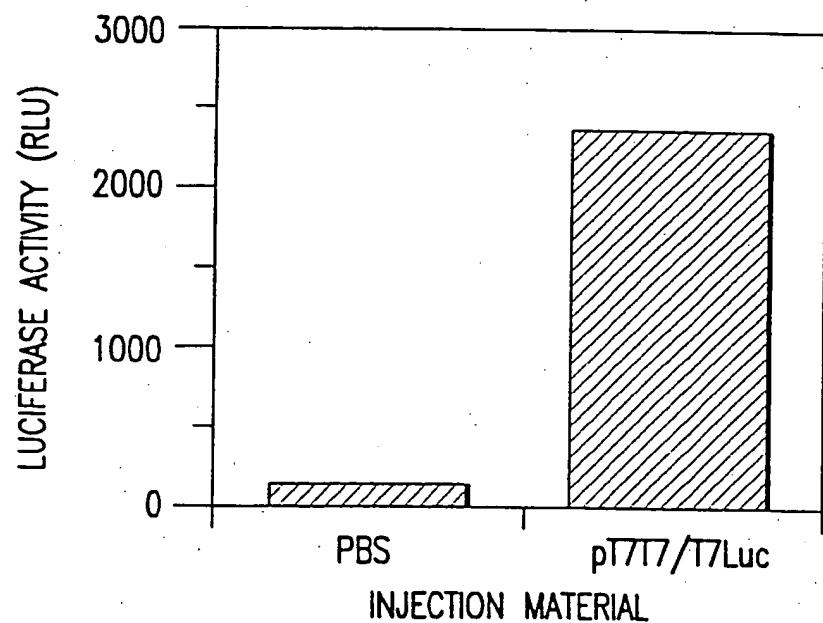


FIG.10

FIG.11
SUBSTITUTE SHEET (RULE 26)

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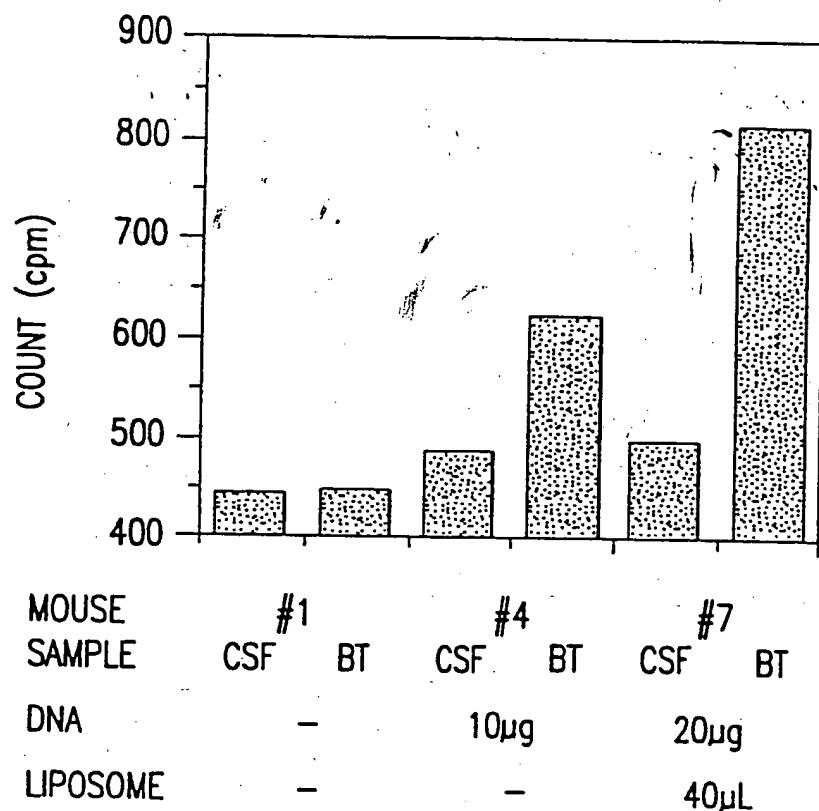


FIG.12

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US94/05409

A. CLASSIFICATION OF SUBJECT MATTER

IPC(5) :C12N 15/63, 15/67, 15/79

US CL :435/69.1, 91.21, 91.31, 172.3, 183, 240.1, 252.3, 320.1

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 435/69.1, 91.21, 91.31, 172.3, 183, 240.1, 252.3, 320.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
none

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, DERWENT BIOTECHNOLOGY ABSTRACTS, CAS, DIALOG, MEDLINE, BIOSIS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	PNAS, Volume 86, issued August 1989, "Elroy-Stein et al., "Cap-independent translation of mRNA conferred by encephalomyocarditis virus 5' sequence improves the performance of the vaccinia virus/bacteriophage T7 hybrid expression system", pages 6126-6130, see entire document.	1-35
Y	Journal of Molecular Biology, Volume 206, issued 1989, Fuerst et al., "Structure and stability of mRNA synthesized by vaccinia virus-encoded bacteriophage T7 RNA polymerase in mammalian cells: Importance of the 5' untranslated leader", pages 333-348, see entire document.	1-35

Further documents are listed in the continuation of Box C. See patent family annex.

* Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"A" document defining the general state of the art which is not considered to be of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
"E" earlier document published on or after the international filing date	"Y"	document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
"L" document which may throw doubt on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	"Z"	document member of the same patent family
"O" document referring to an oral disclosure, use, exhibition or other means		
"P" document published prior to the international filing date but later than the priority date claimed		

Date of the actual completion of the international search	Date of mailing of the international search report
29 JUNE 1994	JUL 20 1994

Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231	Authorized officer JOHN L. LEGUYADER <i>J. Leguyader</i>
Facsimile No. (703) 305-3230	Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US94/05409

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Cell. Biochem., Suppl., 17 Part E, Number S307, issued 1993, Gao et al., "Cytoplasmic gene expression by co-delivery of T7 RNA polymerase and T7 promoter sequence by cationic liposome", page 206, see entire abstract.	1-35
Y	BIOSIS, Abstract No. 96063309, Gao et al., "Cytoplasmic expression of a reporter gene by the co-delivery of T7 RNA polymerase and T7 promoter with cationic liposomes", in Nucleic Acids Research, Volume 21, Number 12, issued 1993, pages 2867-2872, see entire abstract.	1-35